

Robust and Scalable Models of Microbiome Dynamics for Bacteriotherapy Design

Travis E. Gibson¹ Georg K. Gerber^{1,2}

¹Massachusetts Host Microbiome Center
Brigham and Women's Hospital and Harvard Medical School

²Health Sciences and Technology Division Harvard-MIT

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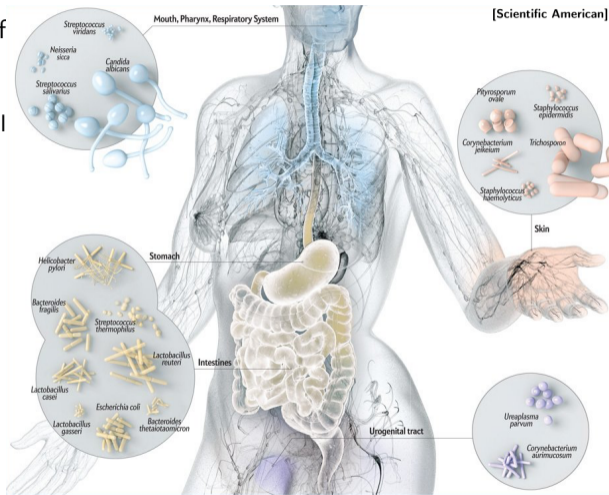
Outline

- ① Background on the Human Microbiome
- ② From Experimental Design to Bacteriotherapies
- ③ Model of microbial dynamics
- ④ Inference Model
- ⑤ Applications

Gerber Lab is looking for Post-docs and PhD students

The Microbiome

- 1 The **microbiome** is the aggregate of microorganisms that resides on or within any of a number of human tissues and biofluids:
 - skin, mammary glands, placenta, seminal fluid, uterus, ovarian follicles, lung, saliva, oral mucosa, conjunctiva, biliary and **gastrointestinal tracts**) [wikipedia]
- 2 10^{14} Microbes in/on your body [Sender et al. *PLoS Biology* 2016]
- 3 3.3 million genes compared to 23,000 human genes [Qin et al. *Nature* 2010]
- 4 Large component of the immune system
- 5 Play a role in a variety of human diseases:
 - infections, arthritis, food allergy, cancer, inflammatory bowel disease, neurological diseases, and obesity/diabetes



Bacteriotherapy

Bacteriotherapy: communities of bacteria administered to patients for specific therapeutic applications

- “bugs-as-drugs”

Clostridium difficile infection

- Causes serious diarrhea (14K deaths/yr)
- Antibiotics disrupt helpful bacteria in gut
- Increasingly difficult to treat with conventional therapies (more antibiotics): 20-30% recurrence rate

Pharmacology meets Ecology



microbial interaction network

positive microbe A produces a small molecule (metabolite) that microbe B needs

negative two microbes competing for the same niche

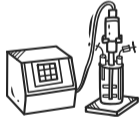
what if there were 300 bugs in the network?

Workflow in our lab

batch
experiments



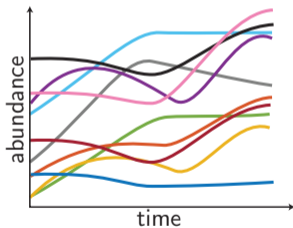
chemostat



animal
experiments



- 16S rRNA on MiSeq
(reads) for relative
abundances of species
- 16S rRNA qPCR
(universal primers) for
bacterial biomass



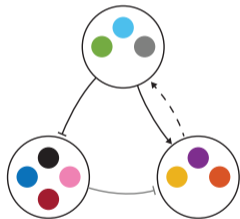
- measurements - irregular,
sparse & noisy

Interaction Network



- 300 species
- 90,000 interactions

Interaction Modules



Microbial Dynamics

- Abundance of microbe i at time t : $\mathbf{x}_{t,i}$

$$\frac{d\mathbf{x}_{t,i}}{dt} = \alpha_i \mathbf{x}_{t,i} + \beta_{ii} \mathbf{x}_{t,i}^2 + \sum_{j \neq i} \beta_{ij} \mathbf{x}_{t,i} \mathbf{x}_{t,j} + \frac{d\mathbf{w}_{t,i}}{dt}$$

growth, carrying capacity, interaction, stochastic disturbance

- Convert to discrete time

$$\mathbf{x}_{k+1,i} = \mathbf{x}_{k,i} + \left(\alpha_i \mathbf{x}_{k,i} + \beta_{ii} \mathbf{x}_{k,i}^2 + \sum_{j \neq i} \beta_{ij} \mathbf{x}_{k,i} \mathbf{x}_{k,j} \right) \Delta_k + (\mathbf{w}_{k+1,i} - \mathbf{w}_{k,i}) \sqrt{\Delta_t}$$

discrete time step size

Next we discuss the three main ingredients to our model

- 1 Clustering (interaction modules)
- 2 Edge selection (structure learning, variable selection)
- 3 Introduction of an auxiliary variable between the measurement model

Complete Model

Dirichlet Process

$$\pi_c \mid \alpha \sim \text{Stick}(\alpha)$$

$$c_i \mid \pi_c \sim \text{Multinomial}(\pi_c)$$

$$b_{c_i, c_j} \mid \sigma_b \sim \text{Normal}(0, \sigma_b^2)$$

Edge Selection (Structure)

$$z_{c_i, c_j} \mid \pi_z \sim \text{Bernoulli}(\pi_z)$$

Self Interactions

$$a_{i,1}, a_{i,2} \mid \sigma_a \sim \text{Normal}(0, \sigma_a^2)$$

Dynamics

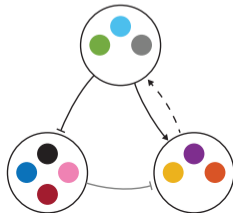
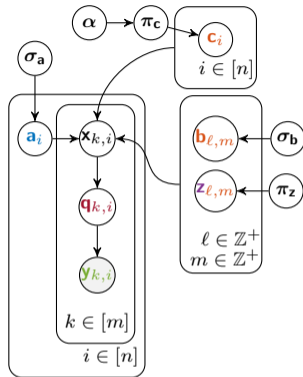
$$\mathbf{x}_{k+1,i} \mid \mathbf{x}_k, \mathbf{a}_i, \mathbf{b}, \mathbf{c}, \mathbf{z}, \sigma_w \sim$$

$$\text{Normal}\left(\mathbf{x}_{k,i} + \mathbf{x}_{k,i} \left(\mathbf{a}_{i,1} + \mathbf{a}_{i,2} \mathbf{x}_{k,i} + \sum_{c_j \neq c_i} b_{c_i, c_j} z_{c_i, c_j} \mathbf{x}_{k,j} \right), \Delta_k \sigma_w^2\right)$$

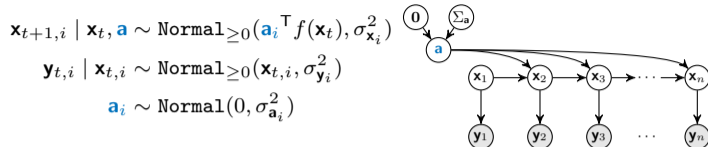
Constraint and Measurement Model

$$\mathbf{q}_{k,i} \mid \mathbf{x}_{k,i} \sim \text{Normal}(\mathbf{x}_{k,i}, \sigma_q^2)$$

$$\mathbf{y}_{k,i} \mid \sigma_y, \mathbf{q}_{k,i} \sim f(\mathbf{q}_{k,i}) \quad f \in \{\text{Neg. Bin.}, \text{Log Norm.}, \dots\}$$



Simple example without the intermediate auxiliary variable



Note the truncated distributions for \mathbf{x} and \mathbf{y}

Parameter inference Gibbs step: $\mathbf{a}^{(g+1)} \sim p_{\mathbf{a}|\mathbf{x}}(\cdot \mid \mathbf{x}^{(g)})$

$$\begin{aligned}
 & \text{Normal}_{\geq 0}(\mathbf{x}; \mu(\mathbf{a}, \mathbf{x}), \sigma^2) \\
 & \uparrow \\
 p_{\mathbf{a}|\mathbf{x}} & \propto p_{\mathbf{x}|\mathbf{a}} p_{\mathbf{x}|\mathbf{a}} p_{\mathbf{a}} p_{\mathbf{a}} \\
 & \downarrow \\
 & \text{Normal}(\mathbf{a}; 0, \sigma^2) \\
 & = \frac{e^{-\frac{1}{2\sigma^2}(\mathbf{x} - \mu(\mathbf{a}, \mathbf{x}))^2}}{\sigma\sqrt{2\pi} \left(\Phi(\infty) - \Phi\left(-\frac{\mu(\mathbf{a}, \mathbf{x})}{\sigma}\right) \right)} \frac{e^{-\frac{1}{2\sigma^2}\mathbf{a}^2}}{\sigma\sqrt{2\pi}}
 \end{aligned}$$

Sampling for other variables

- Filtering (sampling from posterior of \mathbf{x}) is challenging
- Can not use collapsed Gibbs sampling for Dirichlet Process or Edge Selection

Introducing an auxiliary variable

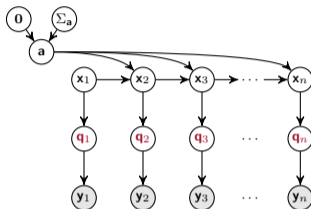
$$\mathbf{x}_{t+1,i} \mid \mathbf{x}_t, \mathbf{a} \sim \text{Normal}(\mathbf{a}_i^\top f(\mathbf{x}_t), \sigma_{\mathbf{x}_i}^2)$$

$$\mathbf{q}_{k,i} \mid \mathbf{x}_{k,i} \sim \text{Normal}(\mathbf{x}_{k,i}, \sigma_{\mathbf{q}}^2)$$

$$\mathbf{q}_{k,i} \sim \text{Uniform}[0, L]$$

$$\mathbf{y}_{k,i} \mid \sigma_{\mathbf{y}}, \mathbf{q}_{k,i} \sim \text{Normal}_{\geq 0}(\mathbf{q}_{k,i}, \sigma_{\mathbf{y}}^2)$$

$$\mathbf{a}_i \sim \text{Normal}(0, \sigma_{\mathbf{a}_i}^2)$$



Prior on \mathbf{q} is positive, relaxing the distribution on the dynamics for \mathbf{x}

Parameter inference Gibbs step: $\mathbf{a}^{(g+1)} \sim p_{\mathbf{a}|\mathbf{x}}(\cdot \mid \mathbf{x}^{(g)})$

- Direct sampling from the posterior now possible (Bayesian Regression!)

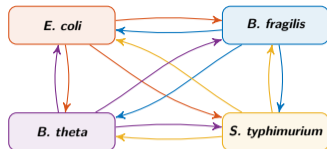
Sampling for other variables

- Collapsed Gibbs sampling for Dirichlet Process and Edge Selection (integrate out \mathbf{a})
- Filtering is still challenging but easier to design proposals than before (MH)

Synthetic consortia of small microbial community

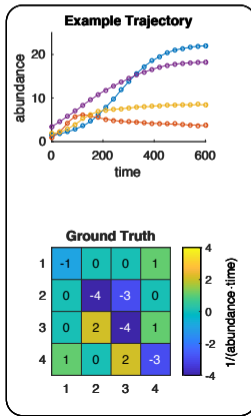


Marika Ziesack
Silver Lab, Harvard

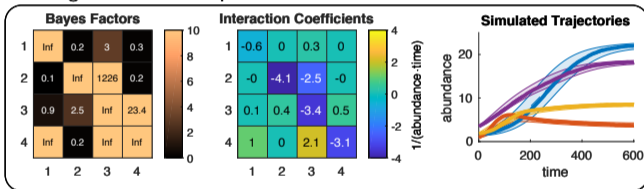


- Microbes engineered to overproduce one amino acid
- Microbes engineered to need three amino acids
- Compare inference on WT and engineered strains to prove that engineering was performed.

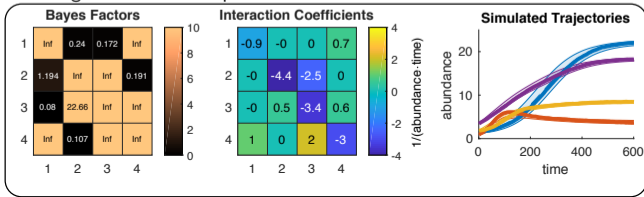
Synthetic Data



Learning from 2 batch experiments

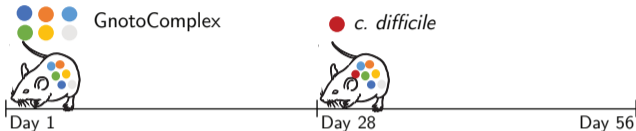


Learning from 4 batch experiments

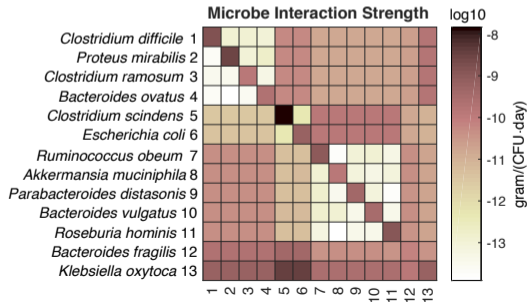
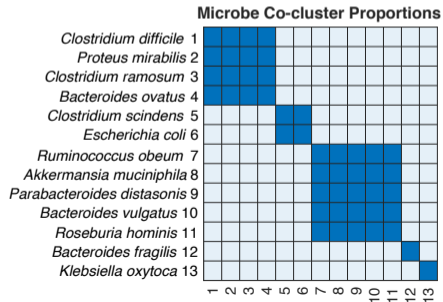


Animal experiments with *Clostridium difficile* infection

- Colonize mice with a defined complex of 12 bacteria (GnotoComplex), then challenge with *Clostridium difficile*



- 5 mice (26 fecal samples taken from each, 16s and universal qPCR)



Conclusions

We have presented

- Fully Bayesian inference model for microbial dynamics
- Interpretability features
 - Reducing the microbial interaction network complexity via extraction of modular features
 - Edge Selection so as to give us confidence as to what interactions are real

Future Directions

- Apply algorithm to mice that have been administered human fecal samples (complex flora 300+ species)
- Approximate Bayesian methods for dynamical systems analysis
- Modeling host dynamics (Layered latent dynamical processes)

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email: tgibson@mit.edu

Gerber Lab Plug

Gerber Lab is looking for post-docs and PhD students

Georg K. Gerber, MD, PhD, (ggerber@bwh.harvard.edu)

- Assistant Professor, Harvard Medical School
- Co-Director, Massachusetts Host-Microbiome Center
- Member of the Harvard-MIT Health Sciences & Technology Faculty
- Associate Pathologist, Center for Advanced Molecular Diagnostics Department of Pathology, Brigham and Women's Hospital